

roboklon

# Blue Pfu DNA Polymerase

## Pfu DNA Polymerase (Pyrococcus furiosus)

 Cat. No.
 Size

 E1116-01
 100 units

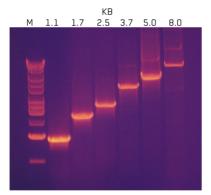
 E1116-02
 500 units

 E1116-03
 2 500 units

Unit Definition: One unit is defined as the amount of enzyme required to catalyze the incorporation of 10 nmoles of dNTP into acid-insoluble material in 30 min at 74°C. The reaction conditions are: 50 mM Tris-HCl (pH 9.0 at 25°C), 50 mM NaCl, 5 mM MgCl<sub>2</sub>, 200  $\mu$ M each of dATP, dCTP, dGTP, dTTP (a mix of unlabeled and I<sup>3</sup>HIdTTP), 10  $\mu$ g activated calf thymus DNA and 0.1 mg/ml BSA in a final volume of 50  $\mu$ l.

## Storage Conditions:

Store at -20°C or at -70 °C.



PCR amplification using EURx Blue Pfu DNA Polymerase. Lane M: molecular size marker - Perfect 1 kb Ladder. Lanes: 1.1 to 3.7 kb: PCR amplification reaction, using buffer B with 0.1 mM dNTPs and 2.5 u EURx Pfu Polymerase in 50  $\mu$ I (0.2 ml tubes) volume, 1 ng lambda DNA as a template, under cycling regime: 94°C 2 min, followed by 35 cycles of 94°C for 20 sec, 55°C for 30 sec and 72°C 1 min/1 kb. Lane 5 kb: PCR as above, using 0.2 mM dNTPs and 3% DMSO. Lane 8 kb: buffer C with 0.35 mM dNTPs, 3% DMSO and 3.75 u EURx Pfu Polymerase. Extremely thermostable proofreading DNA polymerase, suitable for applications requiring high temperature synthesis of DNA. The enzyme is supplemented with an inert blue tracer dye.

## Description:

- → *Pfu* DNA Polymerase is a thermostable enzyme isolated from the hyperthermophilic archaebacterium *Pyroccocus furiosus* (1).
- ➔ Ultrapure recombinant enzyme.
- → Unmodified enzyme replicates DNA at 74°C and exhibits over 95% activity after 1-hour incubation at 98°C.
- → The enzyme catalyzes the polymerization of nucleotides into duplex DNA in the 5' → 3' direction in the presence of magnesium ions.
- → The enzyme also exhibits the  $3' \rightarrow 5'$  proofreading activity, resulting in over 10-fold higher PCR fidelity than possible with *Taq* DNA Polymerases (2).
- ➔ Pfu DNA Polymerase is recommended for use in high-fidelity PCR, PCR of GC-rich sequences or problematic secondary structures, primer extension reactions at elevated temperatures and cloning of blunt-ended PCR products.
- Added blue tracer dye offers several advantages:
  - + visualizes the addition of the polymerase to the reaction;
  - confirms complete mixing;
  - allows for direct loading onto the gel, without the need to add loading buffers and thus saving one step in multiple PCR reactions;
  - + serves as tracking dye during gel electrophoresis;
  - + has no effect on subsequent DNA manipulations such as: ligation, transformation, automated DNA sequencing, restriction digestion;
- → overcomes a frequent problem associated with the red dye, used by other manufacturers, which strongly quenches fluorescence of those DNA bands on the gel, where migration overlaps with the red dye.

## Storage Buffer:

50 mM Tris-HCl (pH 8.2 at 22°C), 0.1 Tween 20, 0.1% Igepal, 0.1 mM EDTA, 1 mM dithiothreitol, 50% glycerol and stabilizers.

#### 10 x Reaction Buffer:

# 10 x AmpliBuffer A (optimization buffer without MgCl<sub>2</sub>):

500 mM KCl, 100 mM Tris-HCl (pH 9.1 at 20°C), 0.1% Triton X-100 and stabilizers. The buffer is optimized for use with 0.1-0.2 mM of each dNTP.

# 10 x AmpliBuffer B (general application, up to 6-8 kb):

500 mM KCl, 100 mM Tris-HCl (pH 9.1 at 20°C), 0.1% Triton X-100, 15 mM MgCl\_ and stabilizers. The buffer is optimized for use with 0.1-0.2 mM of each dNTP.

#### 10 x AmpliBuffer C (for products over 6-8 kb):

160 mM (NH\_4)\_2SO\_4, 500 mM Tris-HCl (pH 9.2 at 22°C), 17.5 mM MgCl\_2 and stabilizers. The buffer is optimized for use with 0.35 mM of each dNTP.

# Quality Control:

All preparations are assayed for contaminating endonuclease, 3'-exonuclease, and nonspecific single- and double-stranded DNase activities. Typical preparations are greater than 95% pure, as judged by SDS polyacrylamide gel electrophoresis.

## References:

- 1. Lundberg, K., Shoemaker, D., Adams, M., Short, J., Sorge, J. and Mathur E. (1991) Gene 108. 1.
- 2. Cline, J., Braham, J. and Hogrefe, H. (1996) Nucleic Acids Res. 24, 3546.

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